Effects of plant extracts and chitosan against Alternaria leaf spot of Chinese kale

Udomsri Ounchokdee and Tida Dethoup*

ABSTRACT

Ethanol extracts of Alpinia galanga (L.)Willd. (rhizome), Coscinium fenestratum (Goetgh.) Colebr. (stem), Piper betle Linn. (leave), Piper longum L. (fruit), Zingiber cassumunar Roxb.(rhizome) and chitosan were evaluated for their fungicidal activities in controlling Alternaria leaf spot of Chinese kale, caused by Alternaria brassicicola, under field conditions. The results showed that the application of these plant extracts resulted in reduction of disease severity in a concentration-dependent manner. Among the five plant extracts tested, the extracts of P. betle and C. fenestratum at a concentration of 10,000 ppm displayed potent fungicidal activity in the suppression of disease severity, causing 43% and 41% disease reduction, respectively. The extracts of Z. cassumunar and A. galanga at 10,000 ppm showed moderate effectiveness in controlling the disease, causing 28% and 25% disease reduction, respectively, whereas the extract of P. longum at the same concentration possessed the lowest antifungal activity, reducing the disease severity by less than 20% at the highest dosed tested. The application of chitosan at 2% had a moderate effect; it reduced the disease severity by 30%. However, the application of the systemic fungicide, iprodione, displayed the greatest fungicidal activity in suppression disease severity, causing 65% disease reduction. The results in this study indicated that the P. betle and C. fenestratum may be promising eco-friendly candidates for controlling Alternaria leaf spot disease in Chinese kale production.

Keywords: Medicinal plant, Phytopathogen control

MS History: 10.02.2020 (Received)-30.04.2020 (Revised)- 16.07.2020 (Accepted)

Citation: Udomsri Ounchokdee and Tida Dethoup. 2020. Effects of plant extracts and chitosan against Alternaria leaf spot of Chinese kale. *Journal of Biopesticides*, **13**(2):159-166.

INTRODUCTION

Alternaria brassicicola is one of the most devastating plant pathogenic fungi. Infection with this fungus results in losses in the yields of host plants. It causes leaf spot disease in wide range of hosts, especially in Brassicaceae including Alternaria blight of rapeseed and mustard (Singh et al., 2017), black leaf spot of cabbage and Chinese kale (Lin et al., 2011; Dethoup et al., 2018), and leaf spot of sugar beet (Rosenzweig et al., 2019). Recently, it was reported as a causal agent of leaf blight on Orychoophragmus violaceus in China (Guo et al., 2019). Under favorable conditions, this plant pathogenic fungus can produce great numbers of spores which infect plant parts and can spread by wind, rain or insects and can infect a crop multiple times (Lin et al., 2011; Siciliano et al., 2017). Moreover, the fungus can infest host plant seeds and can disseminate

as a seed-borne fungus to new crops. The typical symptoms of brown concentric ring on infected leaves are observed within seven days al., after infection (Macioszeket Moreove, this fungus can produce several mycotoxins such as tenuazonic acid, alternariol, alternariol monomethyl ether, altenuene and tentoxin (Pedras and Park, 2015; Siciliano et al., 2017).

JBiopest 13(2): 159-166(2020)

Researchers have been seeking eco-friendly strategies for controlling this pathogen by sprayingcrops with botanical fungicides, plant extracts and biological control agents or treating seed with biological agents, heat or chemicals (Hassan *et al.*, 2017; Dethoup *et al.*, 2018; Rosenzweig *et al.*, 2019; Kokkrua *et al.*, 2020). Plant extracts have been reported to have potent antifungal activity against this pathogen because plants are rich in bioactive compounds (Ounchokdee *et al.*, 2016). The

antifungal activity against this pathogen of extracts of many plant species have been evaluated (Singburaudom, 2015, Rueangrit *et al.*, 2019). Notably, Lin *et al.* (2011) reported the efficacy of *Solanum nigrum* ethanol extract exhibited completely inhibited spore germination of *A. brassicicola* at 500 mg/L whereas the n-butanol fraction of the ethanol extract exhibited 100% spore germination inhibition of *A. brassicicola* at a low concentration of 25 mg/L.

Likewise, chitosan has been reported as a promising biological control agent against many plant diseases (Hadrami et al., 2010). Chitosan is a linear unbranched biopolymer of β-1,4-D- glucosamine derived from a copolymer of D-glucosamine and N-acetyl-Dglucosamine linked with 1,4-glycosidic bonds (Malerba and Cerana, 2018). There are many reports of the potential of chitosan and its mechanisms in controlling plant diseases in many crops (Sharif et al., 2018; Malerba and Cerana, 2018). Multifunctional actions of chitosan in plants have been reported including antifungal and antibacterial activities (Sharif et al., 2018), induced resistance in host plants (Parada et al., 2018), promotion of plant growth (Egusa et al., 2019) and increase in seed germination (Zhou et al., 2017).

Because Chinese kale is an important vegetable in Thailand and because continuous cropping has increased the severity Alternaria leaf spot disease of Chinese kale under field conditions, ecofriendly approaches to control of this disease have been intensively investigated. In our previous study, we evaluated the antifungal activity of 10 plant extracts against this disease and found that crude ethanol extracts of *C. fenestratum* and *P.* betle at 10,000 ppm exhibited significant (P < 0.05) reduction of the disease incidence of up to 67% and showed promising preventive and curative activities against A. brassicicola under greenhouse conditions (Dethoup et al., 2018). However, further studies need to evaluate the antifungal activities of these extracts against this disease under field conditions. In addition, chitosan under various brands has been promoted for controlling this disease in Thailand. Therefore, the objective

of this study was to determine the effects of effective five plant extracts and chitosan at various concentrations in controlling Alternaria leaf spot of Chinese kale under field conditions.

MATERIALS AND METHODS Plant Extractions and Chitosan

The five medicinal plant extracts were prepared as described previously by Dethoup et al. (2018). Briefly, the plants were purchased from a medicinal plant market, in Bangkok, Thailand. Each plant sample was cleaned by washing with tap water thrice, dried in the shade, cut into small pieces, and ground into fine powder with a high-powder grinder. Then, each dried plant preparation was extracted thrice with 70% ethanol at room temperature. The ethanol extract of each plant was filtered through three layers cheesecloth and then concentrated using a rotary evaporator to obtain a crude ethanol extract. The chitosan 2% used in this study was purchased from GEN Technology Co., Ltd., Thailand.

Fungal Pathogen

The strain of *A. brassicicola* KUFA 1079 was isolated from an infected Chinese kale plant, and its pathogenicity was confirmed according to Koch's postulates. The fungus was cultured on potato dextrose agar for 14 days at room temperature. Fifteen mL of sterile water was poured into a petri dish and gently scraped with a sterile glass rod to obtain a spore suspension. The spore suspension was filtered through three layers of sterile cheesecloth and adjusted to 10⁶ spores/ mL using a hemocytometer.

Black spot under field conditions

Field trials were conducted at Amphur Tha Maka, Kanchanaburi province, in the western part of Thailand. Chinese kale seeds were surface disinfected with 0.1% (v/v) sodium hypochlorite for 2 min., washed three times with sterile water, and planted in tray nurseries. The plot size was 1.5 x 1.5 m with 0.1 m within the row and between rows, for a total of 14 rows per plot arranged as a completely randomized design with the distance between plots was 0.5 m.Seven-day-

JBiopest 13(2): 159-166(2020)

161

old seedlings of the same height and vigor were transplanted to plots. Thirty days after planting, the Chinese kale plants were sprayed with one of the following: 1. plant extract at 5,000 ppm; 2. plant extract at 10,000 ppm, 3. chitosan 2%, 4. iprodione 50%WP (1.5 g/L positive control) and 5. water + Tween-20 (negative control). The plants were sprayed with 1 L of each treatment per plot with separate sprayers, and each treatment consisted of three plots (replicates). After 1 hour, the treated plants were inoculated with 1 L of the spore suspension (10⁶ spores/ mL) of

Seven days after applications, thirty leaves were randomly collected from plants in the middle of each test plot. The disease severity was recorded as a percentage of lesion area over the total leaf surface. The disease severity was classified into six levels: 0 = no lesion area; 1 = lesion area 1-20%; 2 = lesion area 21-40%; 3 = lesion area 41-60%; 4 = lesion area 61-80%; 5 = lesion area 41-60%; 4 = lesion area 61-80%; 5 = lesion area 80% (Panwar et al. 2013). The experiments were performed twice during July-August, 2017 and repeated once during August-September, 2018.

Statistical analysis

A. brassicicola per plot.

The experiments in this study were conducted twice. Due to there being no significant difference between the repetitions of each experiment, the separate data of each experiment were pooled and then submitted to analysis of variance, and means were compared by Duncan's multiple range test (*P*

< 0.05), using the statistical program SPSS version 19 (IBM Corporation, Somers, NY).

RERULTS

The effects of five plant crude ethanol extracts chitosan at concentrations Alternaria leaf spot of Chinese kale under field conditions are shown in Table 1 and Fig. 1. The application of the systemic fungicide iprodione showed the greatest fungicidal activity in the suppression of disease severity, causing 68% disease reduction. Among the five plant extracts tested, the extracts of P. betle and C. fenestratum exhibited significant (P < 0.5) reduction of the disease severity. causing 43% and 41% disease reduction, respectively, at a concentration of 10,000 ppm. Following this, the application of chitosan at 2% reduced the disease severity by 30%. The extracts of Z. cassumunar and A. galanga at 10,000 ppm showed moderate antifungal activity in controlling the disease, causing 28.54 and 25.42% disease reduction. respectively, whereas the extract of P. longum displayed the lowest activity, reducing the disease severity by less than 20% at 10,000 ppm. The extracts of C. fenestratum and P. betle at a concentration of 5,000 ppm showed moderate antifungal activity in controlling the disease, causing 26.42 and 29.16% disease reduction, respectively. When applied at 5,000 ppm, the extracts of Z. cassumunar, A. galanga and P. longum exhibited low activity controlling this disease under field conditions.

Table 1 Effects of plant extracts and chitosan on the incidence of Alternaria leaf spot of Chinese kale under field conditions.

Treatment	Plant part	Concentration (ppm)	Disease Severity (%)
Alpinia galanga (L.)Willd.	Rhizome	10,000	$48.32 \pm 4.24^{\rm f}$
		5,000	$59.24 \pm 5.50^{\rm h}$
Coscinium fenestratum (Goetgh.) Colebr.	Stem	10,000	$32.57 \pm 0.89^{\circ}$
		5,000	$47.32 \pm 3.09^{\rm f}$
Piper betle Linn.	Leaves	10,000	30.09 ±2.93 ^b
		5,000	$44.58 \pm 4.41^{\text{de}}$
Piper longum L.	Fruit	10,000	54.27 ± 2.89^{g}
		5,000	61.43 ± 3.62^{i}
Zingiber cassumunar Roxb.	Rhizome	10,000	$45.20 \pm 3.69^{\mathrm{e}}$
		5,000	61.24 ± 6.14^{i}
Chitosan 2%	-	-	43.28 ± 4.58^{d}
Iprodione 50%WP	-	-	5.11 ± 1.04^{a}
Water	-		73.74 ± 6.19^{j}

Means \pm standard derivations followed by the same letter in each row do not significantly differ at P < 0.05, when analyzed using Duncan test of One-Way ANOVA.

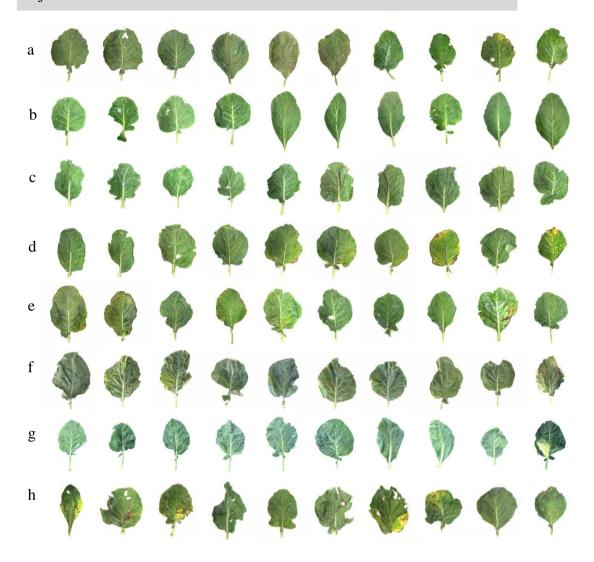


Fig. 1 Effects of plant extracts and chitosan against Alternaria leaf spot disease of Chinese kale under field conditions. a. *Alpinia galanga* extract at 10,000 ppm, b. *Coscinium fenestratum* extract at 10,000 ppm, c. *Piper betle* extract at 10,000 ppm, d. *Piper longum* extract at 10,000 ppm, e. *Zingiber cassumunar* extract at 10,000 ppm f. chitosan at 20,000 ppm, g. iprodione 50% WP (1.5 g/ L) and h. water (negative control)

DISCUSSION

The results in this study showed the efficiency of P. betle and C. fenestratum extracts in controlling Alternaria leaf spot disease of Chinese kale caused by A.brassicicola under field conditions. However, the percentages of disease reduction under field conditions were lower than those of these extracts under greenhouse conditions, which we reported earlier (Dethoup et al., 2018). In our previous report, the applications of C. fenestratum and P. betle extracts were able to reduce the disease severity by up to 67% under greenhouse conditions; however, the extracts reduced the disease severityby43% and 41%, respectively when applied at 10,000 ppm

under field conditions. This phenomenon may be the effect of light, water, wind, and other environmental factors on the performance of the plant extracts against the disease (Deberdt *et al.*, 2008; Rojo Baio *et al.*, 2019). Thus, it may be necessary to increase the concentration of the extracts when applied in field trials due such factors. In contrast, the fungicide, iprodione showed consistent fungicidal effects in controlling the disease under both greenhouse and field conditions (Dethoup *et al.*, 2018).

Plant extracts have been reported as promising agents for controlling plant diseases because the plants from which they are derived produce large groups of bioactive compounds

163

Alternaria leaf spot Management

effective against plant pathogens (Jesonbabu et al., 2012; Dethoup et al., 2019; Kaewsalong et al., 2019). In this study, the extracts of P. betle and C. fenestratum showed potent fungicidal activity against Alternaria leaf spot disease under field conditions. Piper betle, belonging to the family Piperaceae, and its leaf extract been reported to have have antimicrobial activity against many plant pathogens and are also used in folk medicine in many countries (Valentão et al., 2010). Eugenol, β-caryophyllene and hydroxychavicol have been reported as the major bioactive compounds responsible for the antifungal activity of the extract of P. betle leaves (Ali et al., 2010; Valentão et al., 2010; Singburaudom, 2015). There are many reports of the effects of P. betle extract in inhibiting the growth of plant pathogens in vitro including Phaeoisariopsis personata Puccinia arachidis (Kishore and Pande, 2005), Colletotrichum capsici, C. gloeosporioides, Fusarium oxysporium f.sp. cubense Pyricularia oryzae (Singburaudom, 2015).

Meanwhile, Coscinium fenestratum, belonging to the family Menispermaceae, has been found to produce bioactive metabolites against plant diseases. Recently, Kaewsalong et al. (2019) reported the efficacy C. fenestratum extract against dirty panicle disease in rice. The authors found that the extract reduced the disease incidence by 32% when applied at 10,000 ppm under field conditions. Berberine has been reported to be a key bioactive compound in C. fenestratumextract and has also been reported as a main bioactive compound against Alternaria brassicicola, a causal agent of Alternaria leaf spot under greenhouse conditions (Dethoup et al., 2019), and Monilinia fructicola, a causal agent of peach brown rot under field conditions (Fu et al., 2017; Pei et al., 2019). The potency of berberine against plant pathogens results from its many modes of action such as affecting cell wall integrity and ergosterol biosynthesis (Dhamgaye et al., 2014) and binding target DNAs, RNAs and proteins in fungal cells (Fu et al., 2017).

Although the *A. galangal*, *P. longum* and *Z. Cassumunar* extracts showed only moderate

antifungal activity against *A. brassicicola* in this study, there are many studies which reported theses extracts displayed potent antifungal activity against other plant pathogens. *P. longum* extract was reported as an effective agent in inhibiting the mycelial growth of *C. capsici*, *C. gloeosporioides* and *F. oxysporum* f.sp. *cubense in vitro* (Ounchokdee *et al.*, 2016; Rueangrit *et al.*, 2019). Mongkol *et al.* (2014) reported the dichloromethane extract of *A. galangal*

effectively suppressed the growth of *Puccinia nicotianae* and *A. porri* with

MIC31.5 μ g/mL and *C. gloeosporioides* and *F. oxysporum* with MIC250 and 500 μ g/mL, respectively. However, as yet there are no reports about the antifungal activity of these extracts in controlling any plant diseases under field conditions.

Moreover, in this study we also evaluated the efficacy of the commercial chitosan 2% against this disease. Although there are many reports of the potential of chitosan against many plant diseases, it showed moderate fungicide activity against this disease, causing 30% disease reduction. However, Sathiyabama et al. (2014) reported that the application of chitosan at 1 mg/mL could reduce the disease severity of early blight disease of tomato caused by A. solani by 75% while Abd-El-Kareem and Haggag (2014) reported that the application of chitosan at 2 g/L reduced the incidence of early blight of potato caused by A. solani by more than 76.6% and increased tuber yield by more than 80% over the untreated control.

The results in this study indicated that *P. betle* and *C. fenestratum* extracts are promising agents in controlling Alternaria leaf spot of Chinese kale caused by *A. brassicicola* under field conditions. Although the fungicide, iprodione gave the greatest activity in disease reduction, plant extracts are safer and lower in toxicity and negative effects on humans, animals and the environment than that of synthetic fungicides. These findings will support the potential of the extracts of *P. betle* and *C. fenestratum*as alternative approaches for controlling the Alternaria leaf spot disease

in Chinese kale production for sustainable agriculture.

Acknowledgements

This work was financially supported by the Kasetsart University Research and Development Institute under the project "Searching for antifungal compounds from medicinal plants against plant pathogenic fungi for development natural fungicides".

Conflict of Interest

The authors declare no conflict of interest.

References

- Abd-El-Kareem, F. and Haggag, W.M. 2014. Chitosan and citral alone or in combination for controlling early blight disease of potato plants under field conditions. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, **5**:941-949.
- Ali, I., Khan, F.G., Suri, K.A., Gupta, B.D., Satti, N.K., Dutt, P., Afrin, F., Qazi, G.N., and Khan, I.A. 2010. *In vitro* antifungal activity of hydroxychavicol isolated from *Piper betel L. Annals of Clinical Microbiology and Antimicrobials*, **9**:7-9.
- Deberdt, P., Mfegue, C.V., Tondje, P.R., Bon, M.C., Ducamp, M., Hurard, C., Begoude, B.A.D., Ndoumbe-Nkeng, M., Hebbar, P.K., and Cilas, C. 2008. Impact of environmental factors, chemical fungicide and biological control on cacao pod production dynamics and black pod disease (*Phytophthora megakarya*) in Cameroon. *Biological Control*, 44: 149-159.
- Dethoup, T., Songkumorn, P., Rueangrit, S., Suesa-ard, S., and Kaewkrajay, C. 2018. Fungicidal activity of Thai medicinal plant extracts against *Alternaria brassicicola* causing black spot of Chinese kale. *European Journal of Plant Pathology*, **121**: 1-11.
- Dethoup, T., Songkumarn, P., Sirirak, T., and Kijjoa, A. 2019. Fungicidal activity of *Acorus calamus* L. extracts against plant pathogenic fungi. *Agriculture and Natural Resources*, **53**: 526-531.
- Dhamgaye, S., Devaux, F., Vandeputte, P., Khandelwal, N.K., Sanglard, D., Mukhopadhyay, G.,and Prasad, R. 2014. Molecular mechanisms of action of herbal

- antifungal alkaloid berberine, in *Candida* albicans. *PLoS One*, **9**(8):e104554.
- Egusa, M., Parada, R.Y., Aklog, Y.F., Ifuku, S., and Kaminaka, H. 2018. Nanofibrillation enhances the protective effect of crab shells against Fusarium wilt disease in tomato. *Journal of Biological Macromolecules*, **128**: 22-27.
- Fu, W., Tian, G., Pei, Q., Ge, X., and Tian, P. 2017. Evaluation of berberine as a natural compound to inhibit peach brown rot pathogen *Monolinia fracticola*. *Crop Protection*, **91**: 20-26.
- Guo, C., Wang, C.M., Zhou, T.W., Jin, S.L., and Duan C.X. 2019. First report of leaf blight caused by *Alternaria brassicicola* on *Orychophragmus violaceus* in China. *Plant Disease*. doi: 10.1094/PDIS-08-18-1458-PDN
- Hadrami, A.E., Adam, L.R., Hadrami, I.E., and Daayf, F. 2010. Chitosan in plant protection. *Marine Drugs*, **8**: 968-987.
- Hassan, N., Nakasuji, S., Elsharkawy, M.M., Naznin, H.A., Kubota, M., Ketta, H., and Khimizu, M. 2017. Biocontrol potential of an endophytic *Streptomycessp*. Strain MBCN152-1against *Alternaria brassicicola* on cabbage plug seedlings. *Microbes and Environments*, **32**(2): 133-141.
- Jesonbabu, J., Spandana, N., and Aruna Lakshmi, K. 2012. *In vitro* antimicrobial potentialities of chloroform extracts of ethanomedicinal plant against clinically isolated human pathogens. *International Journal of Pharmacy and Pharmaceutical Sciences*, **4**(3): 624-626.
- Kaewsalong, N., Songkumarn, P., Duangmal, K., and Dethoup, T. 2019. Synergistic effects of combinations of novel strains of *Trichoderma* species and *Coscinium fenestratum* extract in controlling rice dirty panicle. *Journal of Plant Pathology*, **101**(2): 367-372.
- Kishore, G.K. and Pande, S. 2005. Integrated management of late leaf spot and rust diseases of groundnut (*Arachis hypogaea* L.) with *Prosopis juliflora* leaf extract and chlorothalonil. *International Journal of Pest Management*, **51**(4): 325-332.

165

Kokkrua, S., Ismail, S.I., Mazlan, N. and Dethoup, T. 2020. Efficacy of berberine in controlling foliar rice diseases. *European Journal of Plant Pathology*, **156**(1): 147-

158.

Alternaria leaf spot Management

- Lin, T.-C., Fan, M.-C., Wang, S.-Y., and Huang, J.-W. 2011. Identification of the *Solanum nigrum* extract component involved in controlling cabbage black leaf spot disease. *Journal of Agricultural and Food Chemistry*, **59**(5): 1667-1672.
- Macioszek, V.K., Lawrence, C.B., and Kononowicz, A.K. 2018. Infection cycle of *Alternaria brassicicola* on *Brassica oleracea* leaves under growth room conditions. *Plant Pathology*, 67(5): 1088-1096.
- Malerba, M., and Cerana, R. 2018.Recent advances of chitosan applications in plants. *Polymers*, **10**(2):118. doi:10.3390/polym10020118
- Mongkol, R., Chavasiri, W., Ishida, M., Matsuda, K., and Morimoto, M. 2014. Phytotoxic and antiphytopathogenic compounds from Thai *Alpinia galangal* (L.)Willd. rhizomes. *Weed Biology and Management*, **15**: 87-93.
- Ounchokdee, U., Rueangrit, S., and Dethoup, T. 2016. Antifungal activity profile of *Piper longum* fruit extract against plant pathogenic fungi. *Journal of Biopesticides*, **9**(2): 97-103.
- Parada, R.Y., Egusa, M., Aklog, Y.F., Miura, C., Ifuku, S., and Kaminaka, H. 2018. Optimization of nanofibrillation degree of chitin for induction of plant disease resistance: elicitor activity and systemic resistance induced by chitin nanofiber in cabbage and strawberry. *International Journal of Biological Macromolecules*, 118: 2185-2192.
- Pedras, M.S.C., and Park, M.R. 2015. Metabolite diversity in the plant pathogen *Alternaria brassicicola*: factors affecting production of brassicicolin A, depudecin, phomapyrone A and other metabolites. *Mycologia*, **107**(6): 1138-1150.
- Pei, Q., Li, Y., Ge, X., and Tien, P. 2019. Multipath effects of berberine on peach

- brown rot fungal *Monilinia fructicola*. *Crop Protection*.**116**: 92-100.
- Rojo Baio, F.H., Antuniassi, U.R., Castilho, B.R., Teodoro, P.E., and da Silva, E.E. 2019. Factors affecting aerial spray drift in the Brazilian Cerrado. *PLoS One*,**14**(2): e0212289.
- Rosenzweig, N., Hanson, L.E., Mambetova, S., Jiang, Q.W., Guza, C., Stewart, J., and Somohano, P. 2019. Fungicide sensitivity monitoring of *Alternaria* spp. causing leaf spot of sugarbeet (*Beta vulgaris*) in the Upper Great Lakes. *Plant Disease*, **103**(9): 2263-2270.
- Rueangrit, S., Eakjamnong, W., and Dethoup, T. 2019. Hidden synergistic effects of the combinations of plant extracts against plant pathogenic fungi. *Journal of Biopesticides*, **12**(1): 51-60.
- Sathiyabama, M., Akila, G., and Charles, R.E. 2014. Chitosan-induced defense responses in tomato plants against early blight disease caused by *Alternaria solani* (Ellis and martin) Sorauer. *Archives of Phytopathology and Plant Protection*, **47**(14): 1777-1787.
- Sharif, R., Mujtaba, M., Rahman, M.U., Shalmani, A., Ahmad, H., Anwar, T., Tianchan, D., and Wang, X. 2018. The multifunctional role of chitosan in horticultural crops; a review. *Molecules*, 23: 872. Doi: 10.3390/molecules23040872.
- Siciliano, I., Gilardi, G., Ortu, G., Gisi, U., Gullino, M.L., and Garibaldi, A. 2017. Identification and characterization of *Alternaria* species causing leaf spot on cabbage, cauliflower, wild and cultivated rocket by using molecular and morphological features and mycotoxin production. *European Journal of Plant Pathology*, **149**(2): 401-413.
- Singburaudom, N. 2015. Hydroxychavicol from Piper betel leave is an antifungal activity against plant pathogenic fungi. *Journal of Biopesticides*, **8**(2): 82-92.
- Singh, H.K., Singh, R.B., Kumar, P., Singh, M., Kumar Yadav, Y., Singh, P.K., Chauhan, M.P., Shakywar, R.C., Maurya, K.N., Shainy Priyanka, B., Srivastciva, T.,

Yadav, S.K., and Maurya, M.K. 2017. Alternaria blight of rapeseed-mustard-a review. *Journal of Environmental Biology*, **38**(60): 1405-1420.

Valentão, P., Gonçalves, R.F., Belo, C., Pinho, P.G., Andrade, P.B., and Ferreres, F. 2010. Improving the knowledge on *Piper betle*: targeted metabolite analysis and effect on acetylcholinesterase. *Journal of Separation Science*, 33: 3168-3176.

Zhou, Y., Jiang, S., Jiao, Y., and Wang, H. 2017. Synergistic effects of nanochitin on inhibition of tobacco root rot disease. *International Journal of Biological Macromolecules*, **99**: 205-212.

Udomsri Ounchokdee and TidaDethoup*

Department of Plant Pathology, Faculty of Agriculture, Kasetsart University, Bangkok 10900, Thailand

*Corresponding author

Tel.: +66 2 579 1026; +66 92 4379991

Fax.: +66 2 579 9550 E-mail: agrtdd@ku.ac.th

ORCID: 0000-0002-9079-3010