

Management of basal rot of onion caused by *Fusarium oxysporum*f. sp. *cepae* using bio-regulators**Rajamohan, K*., Udhayakumar, R., Sanjaygandhi, S., Vengadesh Kumar, L., Thamarai Selvi, M., Sudhasha, S. and Yuvarani, R.****ABSTRACT**

Basal rot of onion caused by *Fusarium oxysporum*f. sp. *cepa* is one of the most serious soil borne diseases causing significant yield loss of up to 50 per cent. Efficacy of various biocontrol agents was evaluated for the potential to manage the basal rot of onion *in vitro*. Among the tested isolates of *Trichoderma viride* (Tv5) gave the greatest (82.86%) inhibition and *Pseudomonas fluorescens* (Pf 2) exerted significantly the greatest (80.82%) reduction of mycelial growth of *F. oxysporum*f. sp. *cepae*. Based on the laboratory analysis, effective biocontrol agents were evaluated in glass house conditions. Among the nine treatments tested in the field by RBD, the combination of bacterial and fungal biocontrol agents and *Glomusmosseae* (Tv5 +Pf2+*G.mosseae*) gave significantly the greatest (89.49%) disease reduction. These biocontrol agents were useful as an alternative to chemical control of the onion basal rot and to enhance growth and yield of onion.

Keywords: Onion, basal rot, *Trichoderma viride*, *Pseudomonas fluorescens*, *Glomusmosseae*

MS History: 30.03.2019 (Received)-20.05.2019 (Revised)- 26.09.2019 (Accepted).

Citation: Rajamohan, K*., Udhayakumar, R., Sanjaygandhi, S., Vengadesh Kumar, L., Thamarai Selvi, M., Sudhasha, S. and Yuvarani, R. 2019. Management of basal rot of onion caused by *Fusarium oxysporum*f. sp. *cepae* using bio-regulators. *Journal of Biopesticides*, **12**(2): 239-247.

INTRODUCTION

Onion (*Allium cepavar. aggregatum*) a bulbous, biennial herb, is one of the most important vegetable crops grown in India (Ilhe *et al.*, 2013). India ranks first in area and second in onion production in the world (Kumar *et al.*, 2015). India has about 1.2 million hectare area under onion cultivation constituting about 10% of total acreage under vegetable with an annual production of 19.40 MT in 2015 (Singh *et al.*, 2017). The total area under production of onion in India during 2016-2017 was 1306 thousand ha with 22427 thousand MT production. However, in Tamil Nadu the total area was 239.286 thousand ha with a total production of 6559.8 thousand MT/ha (NHB 2016). Basal rot of onion caused by *Fusarium oxysporum* f.sp. *cepae* Schlecht. Emend. Snyder and Hansen (Jagatap and suryawnashi, 2015) is one of the most devastating diseases of onion and causes huge yield losses in all growing areas of the world. It causes severe losses in the productivity both

in the field and storage condition (Lager, 2011). Management of this disease through chemicals and the use of resistant varieties are possible to some extent. Bio control is an important component of integrated disease management (IDM) that provides disease control while being relatively harmless to humans, non-polluting and bio-degradable, selective in mode of action, difficult for pathogens to develop resistance, unlikely to harm other beneficial microorganisms and generally improves soil health and sustainability of agriculture and enhances the growth parameters in a number of vegetable crops (Sheo Raj *et al.*, 2004; Jogaiah *et al.*, 2013). Among the biocontrol agents the most commonly used in agriculture are the species of *Trichoderma* which are known to stimulate plant growth by increasing water and mineral uptake, notably that of phosphate, as well as suppressing plant diseases (Shoresh, 2010). The use of *Pseudomonas fluorescens* could provide significant levels of disease

suppression and substantially enhance plant growth and yield. In addition to these antagonists, obligate biotrophic endosymbionts and arbuscular mycorrhizae, are known as plant growth promoting fungi through increasing uptake of mineral nutrients, especially phosphorus (Gill *et al.*, 2002). It is also reported that inoculation of mycorrhizae enhances the growth parameters of onion (Brown *et al.*, 2008). Eco friendly approach will always be better. It ensures maximum suppression of the disease without any adverse effect on the ecosystem. An integrated approach would always ensure greater disease suppression and enhanced crop yield

MATERIALS AND METHODS

Isolation of *F.oxysporumf.sp.cepae*

The pathogen was isolated from the infected bulbs of onion by tissue segment method (Rangaswami, 1958). The infected portions of bulb were cut into small pieces using sterilized scalpel and these were surface sterilized with 1.0 per cent sodium hypo chloride for one minute and washed in three changes of sterile distilled water and then placed on Petri dish containing Potato Dextrose Agar (PDA) medium. These plates were incubated at room temperature ($28 \pm 2^\circ\text{C}$) for five days and observed for the growth of fungus. The hyphal tips of fungi grown from the plates were transferred aseptically to PDA slants for maintenance of the culture. The pathogens were identified based on their cultural and morphological characters.

Isolation of native antagonists from rhizosphere soil *Trichoderma* spp.

Rhizosphere soil samples collected from ten different locations were used for the isolation of *Trichoderma* isolates by soil dilution plating technique using *Trichoderma* selective medium (TSM) (Elad and Chet, 1983). These *Trichoderma* cultures were purified by single hyphal tip method and were used for studies. *Trichoderma* spp., thus isolated was subjected to identification based on the key suggested by Domsch *et al.* (1980). On the whole ten isolates of *Trichoderma* spp. were isolated. These isolates were designated as TV₁-TV₁₀.

Isolation of native antagonistic bacteria

Antagonistic bacteria were isolated from the

rhizosphere soil collected from different onion growing areas of Tamilnadu by serial dilution method on King's B medium, incubating at room temperature for 24 h. Colonies with characteristics of *Pseudomonas* sp., were isolated individually and purified by streaking them on King's B medium. For the identification of *Pseudomonas* sp. isolates, certain biochemical tests were conducted according to Bergey's Manual for Determinative Bacteriology (Breed *et al.*, 1989). The identified isolates were designated as Pf₁-Pf₁₀.

Screening of the fungal and bacterial biocontrol agents against *F.oxysporum f.sp. cepae in vitro*

The antagonistic activity of biocontrol agents against *F.oxysporumf.sp. cepae* was tested by dual culture technique (Dennis and Webster, 1971). At one end of the sterile Petri dish containing 15 ml of sterilized and solidified PDA medium a 9 mm mycelial disc obtained from five day old culture of *Trichoderma* spp. was placed under aseptic conditions. Similarly, at the opposite end approximately 75 mm away from the *Trichoderma* culture disc, a 9 mm culture disc of *F.oxysporumf.sp.cepae* was placed and incubated. Control was maintained by inoculating *F.oxysporumf.sp.cepae* alone at one end of the Petri dish. The plates were incubated at room temperature ($28 \pm 2^\circ\text{C}$) for three days. In case of *P. fluorescens* one cm long streak was gently made onto the medium using two-day-old culture. The radial growth (in mm) of the pathogen and the test antagonists and the extent of the inhibition zones (in mm) developed between the two colonies were measured. The effective antagonists were identified based on the inhibition of the growth of the pathogen. The radial mycelial growth of the pathogen and per cent reduction over control was calculated by using the formula (Vincent, 1927)

$$\text{Per cent inhibition (I)} = \frac{C-T}{C} \times 100$$

Where, C- mycelial growth of pathogen in control

T- Mycelial growth of pathogen in dual plate

I- inhibition per cent

acid and tetramethyl hexadecenol constituted among total identified volatiles.

Efficacy of biocontrol agents against basal rot of onion in pot culture

To study the biocontrol potential of *T. viride*, *P. fluorescens* and *G. mosseae* pot culture experiment was conducted in a glass house. The sand maize culture of the pathogen *F. oxysporum* f. sp. *cepae* were added into 15×30cm diameter earthen pots @ 1.5% W/W of soil. The pots were planted with five bulbs per pot. The talc based formulation of the antagonistics (*Tv5* @ 5g/kg of soil + *Pf2* @ 5g/kg of soil+ *Glomus mosseae* @ 50g/pot) were delivered as basal and soil application at 30 and 45 days after sowing. The pathogen alone inoculated pots served as control and carbendazim (0.1%) was used for comparison. The observations on the per cent disease incidence of basal rot were recorded up to harvest. In addition growth parameters like shoot length, root length and number of bulblets were recorded. Each treatment was replicated thrice.

Statistical analysis

The data collected were subjected to statistical analysis using computer aided IRRISTAT software developed by the International Rice Research Institute, Philippines.

RESULTS AND DISCUSSION

Antifungal activity of *Trichoderma* sp. against mycelial growth of *F.oxysporum* f. sp. *cepae* in *in vitro* (Dual culture technique)

In general all the native *Trichoderma* spp. tested significantly inhibited the mycelial growth of *F.oxysporum* f. sp. *cepae* (Table 1). However, among the isolates, the isolate *Tv5* showed maximum inhibition and significantly inhibited the growth of *F.oxysporum* f. sp. *cepae* (15.42 mm), which was 82.86 per cent reduction on the growth of the pathogen when compared to control. This was followed by the isolates *Tv3* and *Tv9* in the decreasing order of merit, which inhibited the growth of *F.oxysporum* f. sp. *cepae* by 81.98 and 79.86 per cent respectively over control. The least growth inhibition of the pathogen was exhibited by the isolate *Tv7*.

Table 1. Antifungal activity of *Trichoderma viride* against mycelial growth of *F. oxysporum* f. sp. *cepae* in *in vitro* (Dual culture technique)

Isolates	Mycelial growth of <i>F. oxysporum</i> f. sp. <i>cepae</i> (mm)	Per cent (%) inhibition over control
Tv1	35.78 ^g	60.24
Tv2	47.12 ⁱ	47.64
Tv3	16.21 ^b	81.98
Tv4	38.21 ^h	57.54
Tv5	15.42 ^a	82.86
Tv6	23.72 ^e	73.64
Tv7	50.24 ^j	44.17
Tv8	28.82 ^f	67.97
Tv9	18.12 ^c	79.86
Tv10	20.21 ^d	77.54
Control	90.00 ^k	-

*Mean of three replications

*In a column, means followed by a common letter are not significantly different at 5% level by Duncan’s multiple range test (DMRT)

The results of the present study correspond with those of Ilhe (2013), who observed that *T. viride* was found to be most effective in inhibiting the growth of *F. oxysporum* f. sp. *cepae*. Hacer Handan and Oktay (2015) observed that *T. harzianum* 16 and 23 strains showed significant inhibition of mycelial growth of the pathogenic strains of *F. oxysporum*. Both *T. harzianum* strains produced volatile and non-volatile metabolites that inhibited growth of *F. oxysporum* strains. Narayan Prasad Verma *et al.* (2018) showed variation in the antagonistic activities of *Trichoderma* sp. isolates against the tested *Fusarium* sp and also, inhibition of the pathogen may be attributed to the production of secondary metabolites (such as glioviridin, viridin and gliotoxin) the antagonists (Inbar *et al.*, 1994). Several studies (Jayalakshmi *et al.*, 2009; Muhammad and Amusa, 2003; Shabir *et al.*, 2013) reported that inhibition of some soil borne pathogens, including *Fusarium* spp. by

Trichoderma species could probably be due to the secretion of extracellular cell wall degrading enzymes such as chitinase, β -1, 3-glucanase, β -1, 6-glucanase, protease, cellulase and lectin, which help mycoparasites to colonize their host.

Efficacy of native bacterial isolates against *F. oxysporum* sp. *cepae* (Dual culture)

The results presented in Table 2 reveal varying degrees of antagonism by the isolate of *Pseudomonas* against *F. oxysporum* sp. *cepae*. Among the *Pseudomonas* isolates, Pf₂ produced significantly the minimum mycelial growth (23.32 mm) accounting for 80.82 per cent reduction on the mycelial growth of *F. oxysporum* sp. *cepae* over control.

Table 2. Antifungal activity of *Pseudomonas fluorescens* against mycelia growth of *F. oxysporum* sp. *cepae* in *in vitro* (Dual culture technique)

Isolates	Mycelial growth of <i>F. oxysporum</i> sp. <i>cepae</i> (mm)	Percent (%) inhibition over control
Pf ₁	35.73 ^d	60.10
Pf ₂	23.32 ^a	80.82
Pf ₃	48.55 ⁱ	46.05
Pf ₄	29.72 ^b	78.22
Pf ₅	49.62 ^j	44.86
Pf ₆	41.81 ^f	53.54
Pf ₇	31.75 ^c	64.72
Pf ₈	38.98 ^e	56.68
Pf ₉	46.12 ^h	48.75
Pf ₁₀	42.35 ^g	52.94
Control	90.00	-

*Mean of three replications*In a column, means followed by a common letter are not significantly different at 5% level by Duncan's multiple range test (DMRT)

This was followed by isolate Pf₄ which recorded 78.22 per cent reduction on the mycelial growth over control. The isolate Pf₁₀ was the least effective among *Pseudomonas* isolates as it recorded the minimum per cent inhibition control. Boukerma *et al.* (2017) reported that the potential of *P. fluorescens* PF15 and *P.*

putida PP27 showed significant inhibition of the *F. oxysporum* sp. *lycopersici* in tomato. Malathi (2015) reported that Pf 12 and Pf 27 of the *Pseudomonas* isolates were found to be the most effective in inhibiting the growth of *F. oxysporum* sp. *cepae*. Many strains of *Pseudomonas* have been found to produce broad spectrum antibiotics including phenazine, pyrrolnitrin, pyoverdine and 2,4 diacetyl phloroglucinol (Gardener *et al.*, 2000), lytic enzymes such as chitinases and β -1,3-glucanases which degrade fungal chitin (Velazhahan *et al.*, 1999), siderophore (Loper, 1988), HCN (Ahl *et al.*, 1986) and induced systemic resistance (Van Peer *et al.*, 1991). Several earlier workers have suggested a significant role of secondary metabolites such as antibiotics, siderophores of pseudomonads in suppression of fungal pathogens (Vinodkumar *et al.*, 2007; Sreedevi and Charitha Devi, 2012). Such multiplicity of mechanisms exerted by *P. fluorescens* might be the reason for the reduction in the growth of the pathogen.

Effect of combined application of antagonists and *G. mosseae* on basal rot incidence of onion (Pot trial)

The results obtained on the efficacy of combined application of antagonists and *G. mosseae* is furnished in Table 3. Among the treatments, soil application with combination of *T. viride* (Tv₅) and *P. fluorescens* (Pf₂) plus soil application of *G. mosseae* treatment (T₇) recorded the minimum wilt incidence (4.82 %). This was followed by the treatment (T₄) with soil application of *T. viride* (Tv₅) + *P. fluorescens* (Pf₅) which recorded results (7.05 %) at par results with carbendazim 0.1 % in reducing the basal rot incidence. Control recorded the maximum disease (45.87%) incidence. The observations made in the present study are in accordance with the findings of Tayal *et al.* (2011) and Sathiya sivananthamoorthy (2017). Shiva Yendyo *et al.* (2018) also showed that the combined application of native isolates of *Trichoderma* spp. and *P. fluorescens* reduced the disease incidence of *Ralstonia* wilt of tomato.

Table 3. Effect of combined application of antagonists and *G. mosseae* on basal rot incidence of onion (Pot trial)

Tr. No.	Treatments	Basal rot incidence (%)			% disease increase over control		
		30 DAS	45 DAS	At harvest	30 DAS	45 DAS	At harvest
T ₁	SA of <i>Trichoderma viride</i> (5) @ 5g/kg of soil	5.23 ^f	8.21 ^f	9.59 ^f	76.42	72.63	79.15
T ₂	SA of <i>Pseudomonas fluorescens</i> (2)@ 5g/kg of soil	6.52 ^g	9.54 ^g	12.81 ^g	70.60	68.02	72.07
T ₃	SA of <i>Glomus mosseae</i> @ 50g/kg of soil	7.95 ^h	10.67 ^h	15.62 ^h	64.15	64.43	65.94
T ₄	SA of <i>Trichoderma viride</i> (5) @ 5g/kg of soil + SA of <i>Pseudomonas fluorescens</i> (2) @ 5g/kg of soil	2.45 ^b	4.12 ^b	7.05 ^b	88.95	86.26	84.63
T ₅	SA of <i>Trichoderma viride</i> (5) @ 5g/kg of soil + SA of <i>Glomus mosseae</i> @ 50g/kg of soil	4.81 ^d	6.05 ^d	7.12 ^d	85.93	80.06	84.47
T ₆	SA of <i>Pseudomonas fluorescens</i> (2) @ 5g/kg of soil + SA of <i>Glomus mosseae</i> @ 50g/kg of soil	4.52 ^e	7.28 ^e	8.84 ^e	78.31	79.83	80.72
T ₇	SA of <i>Trichoderma viride</i> (5)@ 5g/kg of soil + SA of <i>Pseudomonas fluorescens</i> (2) @ 5g/kg of soil+ SA of <i>Glomus mosseae</i> @ 50g/kg of soil	0.00	2.45 ^a	4.82 ^a	100.00	91.83	89.49
T ₈	Carbendazim 50% WP@0.1%	3.12 ^c	5.82 ^c	7.50 ^c	94.95	87.26	84.63
T ₉	Control	10.00 ⁱ	15.34 ⁱ	45.87 ⁱ	---	---	---

Effect of combined application of antagonists and *G. mosseae* on plant growth promotion of onion (Pot trial)

The efficacy of combined application of antagonists and *G. mosseae* is furnished in Table 4. Among the treatments, soil application with combination of *T. viride* (Tv₅) and *P. fluorescens* (Pf₅₂) and soil application of *G. mosseae* treatment (T₇) recorded the maximum parameters such as shoot length (35.42 cm), root length (5.0 cm) and bulbs (6.0). This was followed by the treatment (T₄) with combination of *T. viride* (Tv₅) and *P.*

fluorescens (Pf₂) which recorded at par results. Control recorded the minimum growth parameters such as shoot length (18.62 cm), root length (2.5 cm) and number of bulbs per plant (1.0). Similar observations were made by Ngullie and Daiho (2013) who recorded reduced incidence of seedling rot in both greenhouse and field condition from the combination of *T. viride* + *P. fluorescens*. Malathi (2015) reported increase in the growth factors like shoot length, root length and number of bulbs with Pf 12 + Pf 27 + TH 3 in the field experiment.

Table 4. Effect of combined application of antagonist and *G. mosseae* on biometrics of onion (Pot trial)

Tr. No.	Treatments	Shoot length (cm)	Root length (cm)	Number of bulbs /plant
T ₁	SA of <i>Trichoderma viride</i> (5) @ 5g/kg of soil	32.52 ^e	4.2 ^c	4.0 ^c
T ₂	SA of <i>Pseudomonas fluorescens</i> (2) @ 5g/kg of soil	30.18 ^f	4.0 ^c	4.0 ^c
T ₃	SA of <i>Glomus mosseae</i> @ 50g/kg of soil	29.15 ^g	3.8 ^f	3.0 ^d
T ₄	SA of <i>Trichoderma viride</i> (5) @ 5g/kg of soil + SA of <i>Pseudomonas fluorescens</i> (2) @ 5g/kg of soil	34.28 ^b	4.5 ^b	5.0 ^b
T ₅	SA of <i>Trichoderma viride</i> (5) @ 5g/kg of soil + SA of <i>Glomus mosseae</i> @ 50g/kg of soil	33.72 ^c	4.2 ^c	4.0 ^c
T ₆	SA of <i>Pseudomonas fluorescens</i> (2) @ 5g/kg of soil + SA of <i>Glomus mosseae</i> @ 50g/kg of soil	32.18 ^d	4.3 ^d	4.0 ^c
T ₇	SA of <i>Trichoderma viride</i> (5) @ 5g/kg of soil + SA of <i>Pseudomonas fluorescens</i> (2) @ 5g/kg of soil + SA of <i>Glomus mosseae</i> @ 50g/kg of soil	35.42 ^a	5.0 ^a	6.0 ^a
T ₈	Carbendazim 50% WP @ 0.1%	32.15 ^e	4.2 ^c	4.0 ^c
T ₉	Control	18.62 ^h	2.5 ^g	1.0 ^e

*Mean of three replications *In a column, means followed by a common letter are not significantly different at 5% level by Duncan's multiple range test (DMRT)

Enhanced plant growth by the siderophore producing strains of fluorescent *Pseudomonas* was reported (Dutta *et al.*, 2005). *P. fluorescens* might have stimulated plant growth by improving uptake of minerals into the host plants particularly phosphate (Kloepper *et al.*, 1980), siderophore mediated iron uptake (Jurkevitch *et al.*, 1988), association with N₂ fixation (Hong *et al.*, 1991), production of IAA (Dubeikovsky *et al.*, 1993), promotion of mycorrhizal function and solubilizing nutrients such as phosphorous. *Trichoderma* spp. are known to produce large quantities of fungistatic metabolites such as trichodermin, dermin, trichoviridin, trichobrachin, chitinase, β -1,3 glucanase, protease etc. (Elad *et al.*, 1982; Bruchkner *et al.*, 1990) which were active against many soil borne pathogens. Thus, the results of the present study and the earlier reports have confirmed that the growth promoting substances produced by *P. fluorescens*, *T. viride* (Tv₃+Pf₅) and *G. mosseae* would have exerted a synergism in promoting the growth parameters of onion.

REFERENCES

- Ahlet, P., Voisard, C. and Defago, G. 1986. Iron bound siderophores, cyanic acid and antibiotics involved in suppression of *Theilaviopsis basicolony* *Pseudomonas fluorescens* strain. *Journal of Phytopathology*. 116 (2): 121-134.
- Boukerma, L., Benchabane, M., Charif, A. and Khelifi, L. 2017. Activity of plant growth promoting rhizobacteria (PGPRs) in the biocontrol of tomato *Fusarium* wilt. *Plant Protection Science*. 53: 78-84.
- Brown, M., Gregon, E., Gapasin, R., Miller, S. and De Castro, A. 2008. Management of soil borne diseases of onions in rice-vegetable system using specific biological control agents (vesicular-arbuscular mycorrhizae, VAM). *Plant Diseases*. 74: 3-27.
- Bruchkner, H., Reinecke, C., Kripp, J. and Kieb, M. 1990. Screening, isolation and sequence determination of a unique group of polypeptide antibiotics from filamentous fungi. *Proc. Int. Mycol. Cong.*

Regensburg, Federation Republic
Germany. P. 224.

- Dubeikovskiy, A. N., Mordukhova, E. A., Kochethov, V. V., Polikarpovo, F. Y. and Boronin, A. M. 1993. Growth promotion of black currant soft wood cuttings by recombinant strain *Pseudomonas fluorescens* BSP53a synthesizing an increased amount of indole-3-acetic acid. *Soil Biology and Biochemistry*. **25**: 1277-1281.
- Dutta, S., Singh, R. P. and Jindal, J. K. 2005. Effect of antagonistic bacteria and plant defense activators on management of bacterial leaf spot of mung bean. *Indian Phytopathology*. **58 (3)**: 269-275.
- Elad, Y., Chet, I., Boyle, P. and Henis, Y. 1982. Parasitism of *Trichoderma* spp. on *Rhizoctoniasolani* and *Sclerotiumrolfsii*-scanning electron microscopy and fluorescent microscopy. *Phytopathology*. **72**: 85-88.
- Dennis, L. and Webster, J. 1971. Antagonistic properties of species-groups of *Trichoderma*. The production of non volatile antibiotics. *Transactions of the British Mycological Society*. **57**: 25-39.
- Elad, Y., Chet, I., Boyle, P. and Henis, Y. 1982. Parasitism of *Trichoderma* spp. on *Rhizoctoniasolani* and *Sclerotiumrolfsii*-scanning electron microscopy and fluorescent microscopy. *Phytopathology*. **72**: 85-88.
- Gill, T. S., Singh, R. S. and Kaur, J. 2002. Comparison of four arbuscular mycorrhizal fungi for root colonization, spore population and plant growth response in chickpea. *Indian Phytopathology*. **55**: 210-213.
- Gardener, B. B. M., Schroeder, K. L., Kaloger, S. E., Raaijmakers, J. M., Thomashow, L. S. and Weller, D. M. 2000. Genotypic and phenotypic diversity of phtD-containing *Pseudomonas* strains isolated from the rhizosphere of wheat. *Applied Environment and Microbiology*. **66 (5)**: 1936-1946.
- Ilhe, M., Musmade, N. A. and Kewade, S. B. 2013. Studies on basal bulb rot of onion caused by *Fusariumoxysporumf. sp. cepae*. *International Journal of Plant Protection*. **6(2)**: 364-366.
- Inbar, J., Abramsky, M. C. D. and Chet, I. 1994. Plant growth enhancement and disease control by *Trichodermaharzianum* in vegetable seedlings grown under commercial conditions. *European Journal of Plant Pathology*. **100**: 337- 346.
- Jayalakshmi, V. and Venkatesan, S. 2006. Effect of biocontrol agents on cassava tuber rot under field conditions. *Indian. Journal of Plant Protection*. **34(1)**: 105-107.
- Jayalakshmi, V. and Venkatesan, S. 2006. Effect of biocontrol agents on cassava tuber rot under field conditions. *Indian. Journal of Plant Protection*. **34(1)**: 105-107.
- Jogaiah, S., Abdelrahman, M., Tran, L. S. P. and Ito, S. 2013. Characterization of rhizosphere fungi that mediate resistance in tomato against bacterial wilt disease. *Journal of Experimental Botany*. **64**: 3829-3842.
- Kumar, V., Neeraj, S. S. and Sagar, N. A. 2015. Post-harvest management of fungal diseases in onion-a review. *Int. International Journal of Current Microbiology and Applied Sciences*. **4 (6)**: 737-752.
- Lager, S. 2011. Survey of *Fusarium* species on yellow onion (*Allium cepa*) on Oland. M.Sc. thesis. Uppsala: Swedish (SLU), Swedish University of Agricultural Science. **PP**. 13-25.
- Loper, J. E. 1988. Role of fluorescent siderophore production in biological control of *Pythiummultimum* by a *Pseudomonas fluorescens* strain. *Phytopathology*, **78**: 166-172.
- Malathi, S. 2015. Biological control of onion basal rot caused by *Fusariumoxysporumf. sp. cepae*. *Asian. Journal of Biological Sciences*. **10 (1)**: 21-26.
- Muhammad, S. and Amusa, N. A. 2003. *In-vitro* inhibition of growth of some seedling

- blight inducing by compost-inhabiting microbes. *African Journal of Biotechnology*. **2** (6): 161-164.
- Narayan Prasad Verma, YugalKishorKuldeep. and Bhisham Kumar Sinha 2018. Efficacy of Indigenous *Trichoderma* Strain Bio-Control against of *Fusarium sp.* Tomato PlantCausal Agent of (*Solanumlycopersicon L.*) *in vitro* Condition. *International Journal of Current Microbiology and Applied Sciences*. **7**(3): 1578-1584.
- Ngullie, M. and Daiho, L. 2013. Efficacy of biocontrol agents in controlling *Rhizoctoniasolaninaga* king Chilli (*Capsicum chinense*Jacq.). *Journal of Experimental Biology and Agricultural Sciences*, **1**: 197-201.
- NHB. Horticultural statistics at a glance. (National Horticultural board), New Delhi (2016).
- Rangaswami, G. 1958. An agar blocks technique for isolating soil micro organisms with special reference to pythiaceous fungi. *Science and Culture* **24**: 85.
- Shabir, R., Rubina, L., Ebenezer, J. K., Talat, M. A., Shabir, A. G., Waseem, A. D. and Javid, A. B. 2013. Eco-friendly management of root-rot of chilli caused by *Rhizoctoniasolanikuehn*. *African Journal of Agricultural Research*. **8**(21): 2563-2566.
- Sheo Raj, Meshram, M. K. and Wasule, D. L. 2004. Emerging biologically based technologies for disease management. In: Khadi BM, Katageri IS, Patil SB,
- Vamadevaiah HM, Udikeri SS, Eshanna editors. Proceedings of the International Symposium on Strategies for Sustainable Cotton Production - A Global Vision. **3**. *Crop Protection* 23-25.
- Shoresh, M., Harman, G. E. and Mastouri, F. 2010. Induced systemic resistance and plant responses to fungal biocontrol agents. *Annual Review of Phytopathology*, **48**: 21-43.
- Singh, M. K., Srivastava, N. and Singh, R. K. 2017. Integrated effect of bio fertilizers and inorganic fertilizers on growth, yield and quality of onion (*Allium cepa*L.). *Journal of Pharmacognosy and Phytochemistry*. **6**(5): 841-1844.
- Sreedevi, B. and Charitha Devi, M. 2012. Mechanism of biocontrol of root rot of groundnut caused by *Macrophominaphaseolina*. *Journal of Agricultural Technology*. **7**(3): 623- 635.
- Jayalakshmi, S. K., Raju, S., Usha, R., Benagi, and Sreeramula, K. 2009. *Trichodermaharzianum*L1 as a potential source for lytic enzymes and elicitor of defense responses in chickpea (*Cicerarietinum*) against wilt disease caused by *Fusariumoxysporum*f.sp. *ciceri*. *Australian Journal of Crop Sciences*. **3**(1): 44-52.
- Jagatap, J. D. and Suryawanshi, N. S. 2015. Potential of biocontrol agents against basal rot of onion caused by *Fusariumoxysporum*f. sp. *cepae*. *International journal of Life Sciences*. **5**: 65-69.
- Jurkevitch, E., Hadar, Y. and Chen, Y. 1988. Involvement of bacterial siderophores in the remedy of lime induced chlorosis in peanut. *Soil Science Society of American Journal*, **52**: 1032-1037.
- Klopper, J. W., Leong, J., Teintze, M. and Schroth, M. M. 1980. Pseudomonas siderophore: A mechanism explaining disease suppressive soils. *Current Microbiology*. **4**: 317-320.
- Hacer Handan Altinok. and OktayErdogan. 2015. Determination of the *In vitro* Effect of *Trichodermaharzianum* on Phytopathogenic Strains of *Fusariumoxysporum*. *Nat. Bot. Horti. Agro. bo*. **43**(2): 494-500.
- Hong, Y., Pasternak, J. J. and Glick, B. R. 1991. Biological consequences of plasmid transformation of the plant growth promoting rhizobacterium *Pseudomonas*

putida GR12-2. *Canadian Journal of Microbiology*. **37**: 796-799.

Velazhahan, R., Samiyappan, R. and Vidhyasekaran, P. 1999. Relationship between antagonistic activities of *Pseudomonas fluorescens* isolates against *Rhizoctonia solani* and their production of lytic enzyme. *Journal of Plant Diseases and Protection*. **106** (3): 244-250.

Van Peer, R., Niemann, G. J. and Schippers, B. 1991. Induced resistance and phytoalexin accumulation in biological control of *Fusarium* wilt of carnation by *Pseudomonas* spp. strain WCS 417K. *Phytopathology*, **81**: 728-734. Vinodkumar A, Verma VC, Gund SK and Kharwar RN (2007). Induction of systemic resistance by plant growth promoting

247
rhizobacteria against red rot disease caused by *Colletotrichum falcatum* Went. In sugarcane. *Proc. Sugar Technol. Assoc. India*. **61**: 24-39.

Rajamohan, K¹, Udhayakumar, R., Sanjaygandhi, S., Vengadesh Kumar, L., Thamarai Selvi, M., Sudhasha, S. and Yuvarani, R.

Department of Plant Pathology, Annamalai University, Annamalainagar – 608002, Tamil Nadu, India.

*Corresponding author

E-mail: rajamohan.pat@gmail.com